

Analysis of Presatovir (GS-5806) Resistance Emergence in Human Healthy Adult Subjects Experimentally Infected With Respiratory Syncytial Virus

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Introduction

- Presatovir is a potent, small-molecule respiratory syncytial virus (RSV) fusion inhibitor with activity against a broad spectrum of RSV A and B clinical isolates
- The efficacy of presatovir was evaluated in a randomized, double-blind, placebo-controlled, Phase 2a viral challenge study in healthy adults (aged 18–45 years) experimentally infected with RSV Memphis 37b (NCT01756482)¹

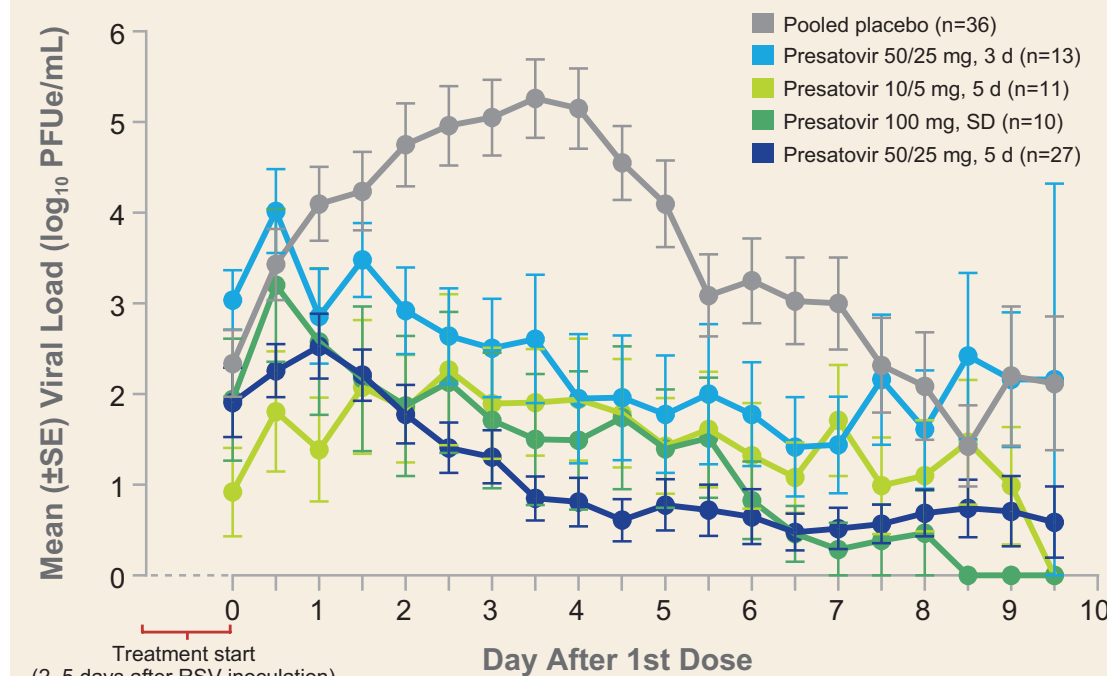
Presatovir Challenge Study Design*

Subjects/Cohort	Presatovir	Placebo	Cohort Presatovir mg/d or placebo				
			1–4 50/25 mg, 5 d	5 50/25 mg, 3 d	6 100 mg, SD	7 10/5 mg, 5 d	
39	39		50	25	25	25	25
17	5		50	25	25		
14	4		100				
17	5		10	5	5	5	5

*Subjects were randomized into 7 sequential treatment cohorts. Subjects with low serum RSV neutralizing antibody titers were inoculated via intranasal administration with ~4 log₁₀ plaque-forming unit equivalents (PFUe) of RSV Memphis 37b. Subjects were monitored for RSV in nasal wash samples by qualitative reverse-transcription-polymerase chain reaction (RT-PCR) assay and were randomized into cohorts when RSV-positive or 5 days after inoculation. SD, single dose.

- In all cohorts, presatovir treatment reduced viral load from nasal wash samples, total mucus weight, and symptom scores compared with placebo

Posttreatment Viral Load*



*Viral load measured by quantitative RT-PCR assay was plotted from start of treatment to end of quarantine period (Day 12). SE, standard error.

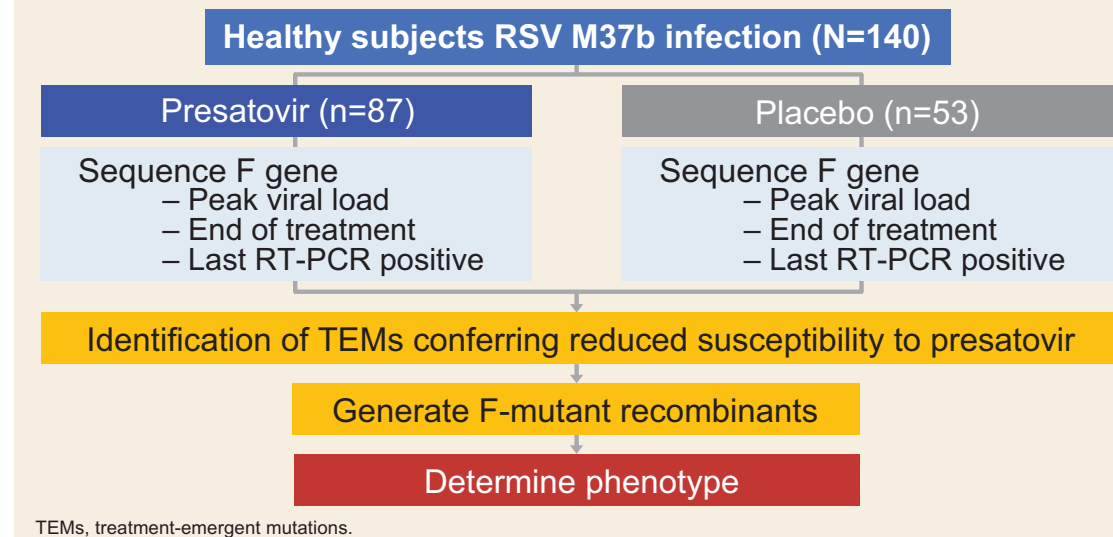
- No significant adverse events were reported
- Presatovir is currently being evaluated in several Phase 2a studies in adults with natural RSV infection (NCT02254421, NCT02254408, NCT02534350, and NCT02135614)
- Here we present the results of drug-resistance emergence analyses in subjects enrolled in the Phase 2a RSV challenge study

Objectives

- To identify and characterize treatment-emergent presatovir-resistant RSV variants

Methods

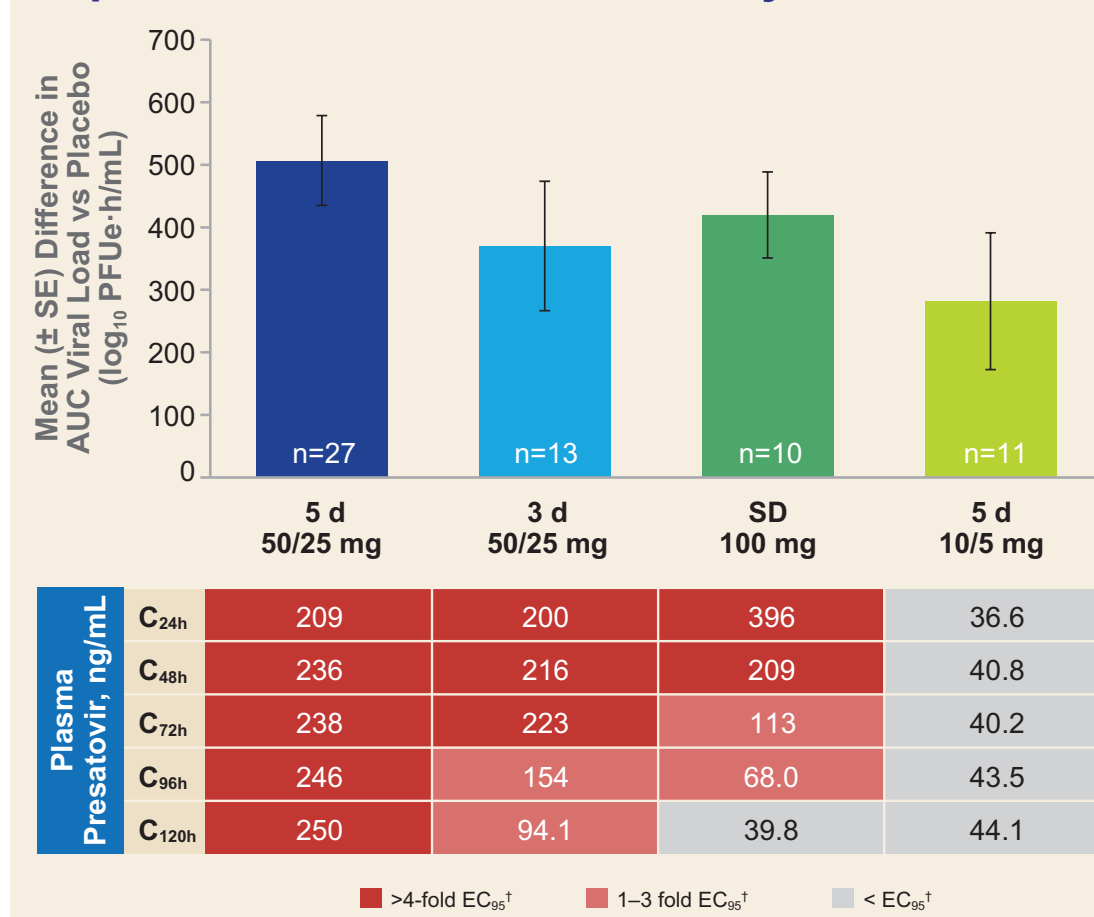
Resistance Sampling



- RSV sampling: emergence of drug resistance analyzed by population sequencing of RSV from nasal wash samples taken at viral load peak, end of dosing, and last day of RSV detection
- RSV F-gene sequencing
 - Viral RNA extracted from nasal wash samples using SV 96 Total RNA Isolation System (Promega Corporation, Madison, WI)
 - RSV F gene amplified by QIAGEN OneStep RT-PCR Kit (QIAGEN, Valencia, CA)
 - Nucleotide sequence of RSV F gene determined by Sanger sequencing
- RSV recombinant virus production
 - Point mutations representing individual TEMs introduced by site-directed mutagenesis into plasmid clones encoding wild-type (WT) RSV F protein
 - Recombinant mutant RSV strains expressing individual TEMs constructed by transferring mutant RSV F gene into RSV genome using modified RSV reverse-genetics methodology²
- Phenotypic analysis
 - Susceptibility of RSV recombinants containing TEMs related to RSV Memphis 37b determined in RSV cell-based antiviral enzyme-linked immunosorbent assay (ELISA)
 - HEp-2 cells (3x10³ cells/well) infected with RSV 100 µL (1x10^{4.5} tissue infectious doses/mL) in media in presence of serial dilutions of presatovir
 - Virus replication quantified 4 days postinfection by RSV-specific ELISA using RSV F-protein-specific murine monoclonal antibody
- Competitive fitness assay
 - Mixtures of WT and TEM-expressing RSV passaged 3 times in HEp-2 cells
 - Each virus contained a silent-marker sequence in the N gene that was recognized by unique TaqMan[®] probes
 - The proportions of WT and presatovir-resistant variants were determined by a multiplexed quantitative RT-PCR assay using probe pairs with minimal spectral overlap to quantify the level of WT or resistant variants

Results

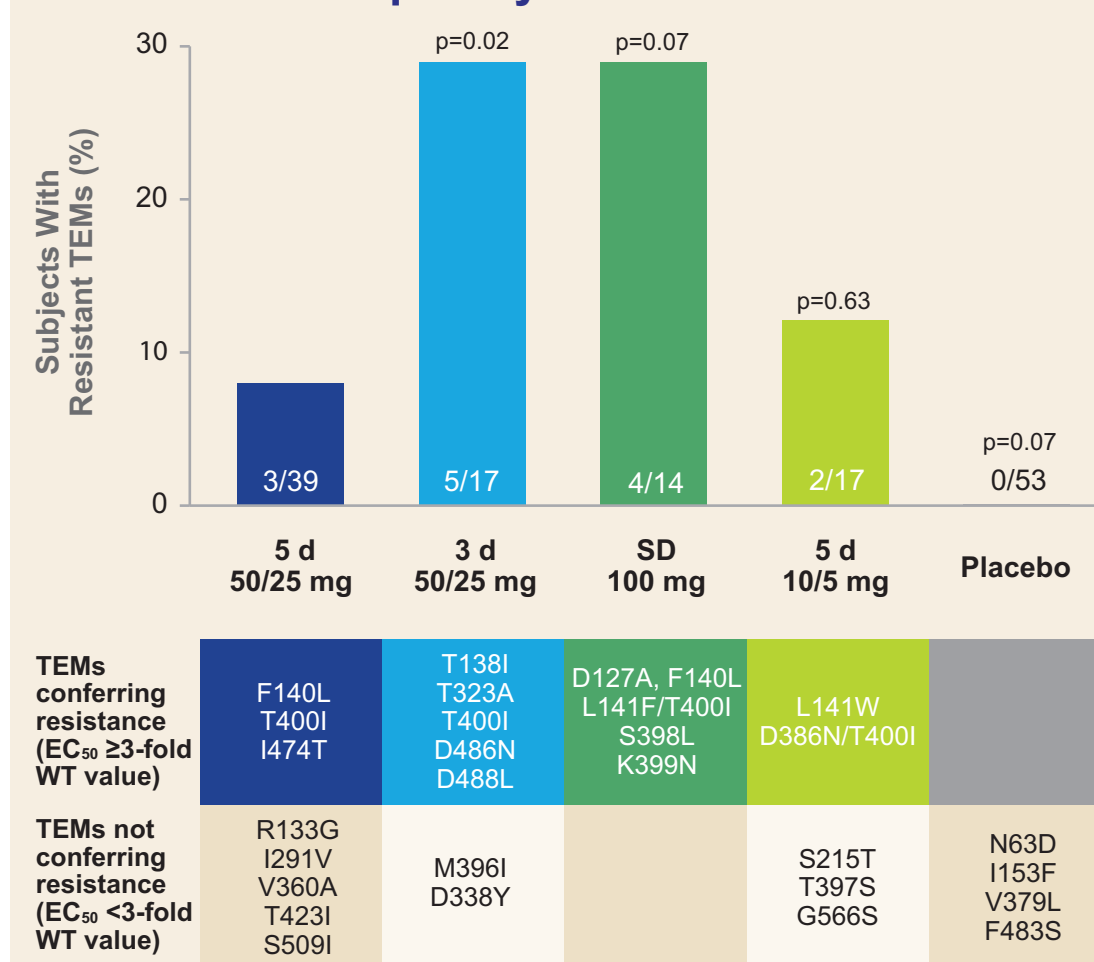
Exposure-Driven Antiviral Activity of Presatovir*



*Area under curve (AUC) viral load analysis restricted to subjects who were RSV positive within 5 days postinoculation; [†]95% maximal effective concentration (EC₉₅) for RSV Memphis 37b. C, concentration.

- Exposure level of presatovir above EC₉₅ for RSV Memphis 37b correlated with antiviral response

Resistance Frequency*



*Calculated by dividing number of subjects with resistant variants by total number of presatovir-treated subjects. p-values for each cohort are relative to 5d (50/25 mg) cohort. EC₅₀, half maximal effective concentration.

Antiviral Susceptibility of RSV Variants Expressing TEMs Associated With Presatovir Resistance*

Resistant variant	Presatovir	Palivizumab [†]	Ribavirin
	EC ₅₀ shift relative to RSV A2		
WT RSV A2 EC ₅₀ , µM	0.0002 ± 0.0001	0.23 ± 0.27	12.41 ± 5.04
L138I	>200	0.7	0.8
V127A, F140L [‡]	>200	0.1	0.7
F140I	>200	0.9	1.5
L141F	>200	0.6	0.7
L141W	>200	1.5	1.2
L141F/T400I [§]	>200	0.5	1.2
T323A	5.0	0.5	0.5
D338Y	2.9	0.8	0.8
S398L	>200	0.3	0.3
K399N	38	0.6	0.6
T400I	>200	1.3	1.2
T400V	>200	1.5	1.2
T400A	>200	3.3	1.8
I474T	3.2	0.7	1.2
D486N	>200	0.8	1.3

*Data represent mean of ≥3 experiments; [†]Values in µg/mL; [‡]Mutants genetically linked; [§]Mutants not genetically linked.

Conclusions

- Treatment-emergent mutations (TEMs) conferring reduced susceptibility to presatovir were identified in the F gene of several viral isolates from subjects experimentally challenged with RSV and treated with presatovir
- The frequency of resistance to presatovir was lowest for the 5 day 50/25-mg treatment group
- Subjects harboring presatovir-resistant variants had higher AUC viral load values than subjects with WT virus
- Clinical significance of TEMs is unknown: no significant differences were observed in clinical symptom scores or mucous weights between subjects with presatovir-resistant variants and WT virus
- RSV recombinants with reduced susceptibility to presatovir remained sensitive to inhibition by palivizumab and ribavirin
- The most frequently observed presatovir-resistant TEMs (F140L and T400I) conferred reduced in vitro replicative fitness of RSV
 - In vivo assessment of replication capacity is in progress

References

1. Devincenzo J, et al. *N Engl J Med* 2014;71:711-22; 2. Collins PL, et al. *Proc Natl Acad Sci USA* 1995;92:11563-7.

Acknowledgments

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