

# Determinants for Antibiogram Development for use by Skilled Nursing Facilities in a Large Geographic Region, 2016-2017

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## Background

- Skilled nursing facilities (SNFs) are an important and growing component of the U.S. healthcare system and recognized as settings in which resistant pathogens and inappropriate antimicrobial use are prevalent (1-2).
- As of January 2018 all SNFs are required to have an antibiotic stewardship program (ASP) (3-4), including access to cumulative susceptibility data (antibiograms) from isolates recovered from infections among residents (5).
- However, most SNFs submit so few specimens in a given year that constructing SNF-specific antibiograms for a single year cannot be performed consistent with CLSI Guideline M39-A4 (30>= isolates per organism recommended (6,7,8).
- Sharing data across SNFs within a given year is one method to overcome this limitation of data and provide more robust antibiograms for each SNF in a region.

## Objectives

This analysis explores best methods to combine data across SNFs in order to enable clinical laboratory staff producing antibiograms to assess:

- Are there significant regional differences in AR (antibiotic resistance) patterns?
- Should groups of SNFs be combined to produce an antibiogram based on SNF characteristics?

## Methods

- Retrospective observational cohort study of SNF residents in 2016-2017 with urine culture processed by Clinical Laboratory Services (CLS) in Winder, GA.
- Testing results in study population were limited to:
  - CMS (Centers for Medicare and Medicaid Services) designated skilled nursing facilities in the GA
  - E. coli*, *K. pneumoniae*, *P. aeruginosa*, *P. mirabilis*
  - Patients aged 65-75 years.
- Susceptibility testing was performed using a MicroScan Walkway96 Plus, select results were suppressed consistent with standards (2nd gen suppressed if gram negative fermenters test susceptible to cefazolin)
- Linkage by National Provider Number to a CMS Certification Number (CMS CCN) allowed abstracting facility characteristics from 2016 Nursing Home Compare
- Log-Binomial Regression was performed at the isolate level to identify critical facility or encounter characteristics predictive of testing susceptible.
- Spider plots of estimated % susceptible were plotted by characteristic to evaluate consistent differences in % susceptible across antibiotic-pathogen pairs.
- Acronyms: CEF3 = non-susceptible to any 3<sup>rd</sup> generation cephalosporin tested, FQ = non-susceptible to either ciprofloxacin or levofloxacin.

Table 1. No. of facilities submitting isolates, and no. of isolates per facility, by pathogen.

Organism	All Facilities		No. of Isolates per Facility					
	No. of facilities	% submitting >1 isolate	Mean	Min	25%	50%	75%	Max
<i>E. coli</i>	195	74	6.25	1	2	4	8	35
<i>K. pneumoniae</i>	145	55	3.14	1	1	2	4	14
<i>P. aeruginosa</i>	73	59	2.34	1	1	2	3	16
<i>Proteus mirabilis</i>	154	28	3.25	1	1	2	4	17
<i>E. faecium</i>	50	19	1.40	1	1	1	1	7

- Crude values will be heavily weighted by facilities reporting more pathogens

Table 2. Estimated Percent Susceptible for Select Urinary Pathogens, by geographic region.

- Yellow boxes illustrate statistically significant differences in %S between two regions, orange boxes are of borderline statistical significance. Overall only 18 of 144 comparisons were statistically significant (10 of these 18 involved the northern region). HD3 = Health District 3.

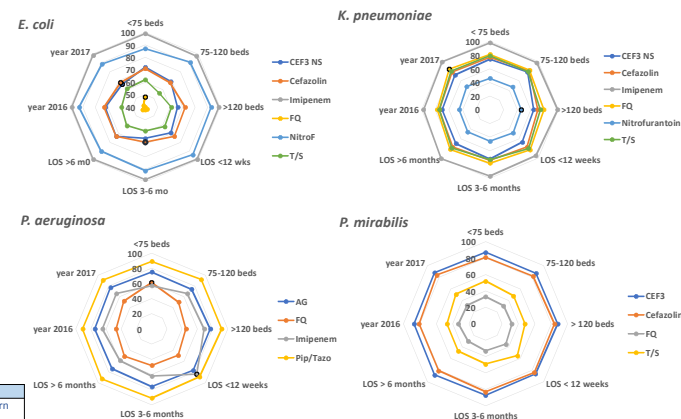


Pathogen	Antibiotic	% Susceptible, by Region				Mean difference in percentage points comparing two regions (P-value)							
		Central	HD3	North	South	Central to Atlanta HD3	Central to Southern	Atlanta HD3 to Southern	Northern to Central	Northern to Atlanta HD3	Northern to Southern	Northern to Southern	
<i>E. coli</i>	CEF3	73.1	63.6	62.1	73.8	9.5 (0.01)	-0.7 (0.88)	-10.1 (0.03)	11 (0.01)	1.5 (0.74)	-12 (0.02)	-13 (0.06)	
	Cefazolin	71.8	69.7	65.5	73.5	2.1 (0.58)	-1.7 (0.70)	-3.8 (0.41)	6.4 (0.14)	4.3 (0.35)	-8.0 (0.11)	-8.0 (0.11)	
	FQ	42.9	41.7	32.9	46.5	1.3 (0.74)	-3.7 (0.40)	-4.9 (0.29)	10 (0.02)	8.8 (0.05)	-14 (0.01)	-14 (0.01)	
	Imipenem	99.2	99.0	97.3	98.3	0.3 (0.95)	0.9 (0.83)	0.7 (0.88)	2.0 (0.65)	1.7 (0.71)	-1.0 (0.84)	-1.0 (0.84)	
	Nitrofurantoin	91.0	92.1	89.0	89.7	-1.1 (0.77)	1.2 (0.77)	2.4 (0.61)	2.0 (0.64)	3.1 (0.50)	-0.7 (0.88)	-0.7 (0.88)	
<i>K. pneumoniae</i>	Trimeth/Sulfa	59.4	62.6	53.7	62.2	-3.2 (0.40)	-2.8 (0.51)	0.4 (0.93)	5.7 (0.18)	8.9 (0.05)	-8.5 (0.09)	-8.5 (0.09)	
	CEF3	64.5	65.7	82.7	78.7	-1.2 (0.82)	-14 (0.02)	-13 (0.04)	-18 (0.01)	-17 (0.01)	4.0 (0.56)	4.0 (0.56)	
	Cefazolin	71.1	72.9	86.2	78.6	-1.7 (0.74)	-7.5 (0.21)	-5.8 (0.35)	-15 (0.01)	-13 (0.03)	7.6 (0.27)	7.6 (0.27)	
	FQ	76.3	77.0	87.6	86.3	-0.6 (0.91)	-10 (0.10)	-9.4 (0.13)	-11 (0.07)	-10 (0.09)	1.2 (0.86)	1.2 (0.86)	
	Imipenem	98.9	97.4	97.8	97.0	1.4 (0.79)	1.9 (0.75)	0.5 (0.94)	1.1 (0.86)	-0.4 (0.95)	0.8 (0.91)	0.8 (0.91)	
<i>P. aeruginosa</i>	Nitrofurantoin	46.5	45.6	53.7	40.7	0.8 (0.87)	5.8 (0.34)	4.9 (0.43)	-7.2 (0.24)	-8.1 (0.20)	13 (0.06)	13 (0.06)	
	Trimeth/Sulfa	73.6	73.0	84.0	81.0	0.6 (0.90)	-7.3 (0.22)	-8.0 (0.19)	-10 (0.09)	-11 (0.08)	3.0 (0.66)	3.0 (0.66)	
	AG	78.2	72.2	84.0	73.4	6.0 (0.44)	4.8 (0.60)	-1.2 (0.90)	-5.8 (0.58)	-11 (0.24)	-10 (0.34)	-10 (0.34)	
<i>P. mirabilis</i>	FQ	46.7	42.5	63.6	57.5	4.2 (0.58)	-10 (0.24)	-15 (0.09)	-16 (0.11)	-21 (0.03)	6.1 (0.59)	6.1 (0.59)	
	Imipenem	62.3	67.6	66.5	65.4	-5.3 (0.49)	-3.1 (0.74)	2.2 (0.80)	-4.2 (0.69)	1.1 (0.91)	1.1 (0.92)	1.1 (0.92)	
	Pip/Tazo	93.5	89.0	92.3	90.3	4.4 (0.56)	3.2 (0.73)	-1.3 (0.88)	1.1 (0.91)	-3.3 (0.74)	2.1 (0.85)	2.1 (0.85)	
	CEF3	87.1	87.3	88.7	85.7	-0.2 (0.98)	1.5 (0.80)	1.6 (0.79)	-1.6 (0.78)	-1.4 (0.81)	3.0 (0.65)	3.0 (0.65)	
<i>P. mirabilis</i>	Cefazolin	83.0	84.9	86.1	76.0	-1.9 (0.70)	7.0 (0.23)	8.9 (0.14)	-3.1 (0.59)	-1.2 (0.85)	10 (0.13)	10 (0.13)	
	FQ	25.1	33.6	30.5	44.7	-8.9 (0.09)	-19 (0.00)	-11 (0.07)	-5.4 (0.35)	3.2 (0.60)	-14 (0.03)	-14 (0.03)	
	Trimeth/Sulfa	50.5	46.4	43.8	55.4	4.1 (0.41)	-4.8 (0.41)	-8.9 (0.14)	6.8 (0.24)	2.7 (0.65)	-11 (0.09)	-11 (0.09)	
	No. of comparisons (%) with significant/borderline differences	2 (9%)	2 (9%)	3 (13%)	8 (33%)	9 (38%)	7 (29%)						

- Significant differences in %S by geographic region was rare, occurring in only 18 of 144 possible comparisons; however, the majority of differences involved the northern region (Table 2)

## Results

- Figure 1. Spider plots illustrating the percent of isolates (one pathogen per figure) testing susceptible to each drug (designated by different colors).
- The vertical axis is estimated % susceptible by the statistical model adjusting for characteristic on an axis of "spider."
- Black circles indicate characteristic significantly higher or lower %S.
- No differences would result in a symmetrical octagon.



- Any single SNF characteristics were not consistently associated with different % susceptibility rates; although average LOS (Length of Stay), bed size, or year affected some drug-bug combination rates (Figure 1).
- These data are limited in age and time, the observed differences may become significant with additional data added to the analysis.

## Conclusions

- SNFs submit too few specimens to produce a SNF-specific antibiograms;
- Combining test results across SNFs is an attractive solution to produce viable reports. There appears to be little statistical justification to producing distinct antibiograms for a SNF by grouping "similar" SNFs based on a SNF characteristic (e.g., beds). Likewise, in GA, a regional antibiogram could be produced for northern vs. other areas, but the observed differences were still of little clinical relevance.

## Contact

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## References

- Centers for Disease Control and Prevention. Core Elements of Hospital Antibiotic Stewardship Programs. Atlanta, GA: US Department of Health and Human Services, CDC; 2014 Accessed 9/30/2014.
- Lim CJ, Kong DCM, Stuart RL. Reducing inappropriate antibiotic prescribing in the residential care setting: current perspectives. Clin Interv Aging. 2014; 9: 165-177.
- Furuno JP, Comer AC, Johnson JK, et al. Using antibiograms to improve antibiotic prescribing in skilled nursing facilities. Infect Control Hosp Epidemiol. 2014;35 (Suppl 3):S56-61.
- Dellit TH, Owens RC, McGowan JE, Jr., et al. Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America guidelines for developing an institutional program to enhance antimicrobial stewardship. Clinical infectious diseases. 2007;44(2):159-177.
- Jump RLP, Olds DM, Seifi N et al. Effective antimicrobial stewardship in a long-term care facility through an infectious disease consultation service: Keeping a lid on antibiotic use. Infect Control Hosp Epidemiol 2012;33(12):1185-1192.
- Nicolle LE. Antimicrobial stewardship in long-term care facilities: what is effective? Antimicrob Resist Infect Contr 2014; 3:6. Accessed 12/3/14.
- Ray, G.T., J.A. Suaya, and R. Baxter. Trends and characteristics of culture-confirmed *Staphylococcus aureus* infections in a large U.S. integrated health care organization. J Clin Microbiol. 2012. 50(6): p. 1950-7.
- Clinical and Laboratory Standards Institute. Analysis and Presentation of Cumulative Antimicrobial Susceptibility Test Data; Approved Guideline. Fourth Ed. M39-A4. 01/05/2014. ISBN Number: 1-56238-899-1